INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACY AND CHEMISTRY

Available online at www.ijrpc.com

Research Article

DOI: https://dx.doi.org/10.33289/IJRPC.9.4.2019.958

ANTI-CANCEROUS ACTIVITY OF AUTO-OXIDIZED PRODUCTS

FROM 9H, 10H-INDOLIZINO [1, 2-B] INDOLE-1-ONE DERIVATIVES

Gautam Bhattacharya

Department of Chemistry, Maharaja Udaychand Women's College, B.C. Road, Burdwan, 713 104, West Bengal, India.

ABSTRACT

The tetracycle 9H, 10H-Indolizino [1, 2-b] indole-1-one derivatives (2a - 2d) underwent a novel auto-oxidation in the presence or absence of base ¹. The product peroxides (7a - d, 10a - d, 12a - d) were tested for their anti-cancerous activity against different cancer cell lines. Few peroxides were found out to be active against cancer cell lines with IC₅₀ in low nanomolar to macromolar range. Our study proves the efficacy of the peroxide to stop propagation of cancerous cell lines *in vitro*.

Keywords: Auto-oxidation, indolizino indole, peroxides, in vitro anti-cancer activity and radicals.

INTRODUCTION

Development of new anti-cancerous drugs is the call of the hour, as cancer is becoming an almost epidemic in modern time, may be due to our lifestyle and modernization of society. In our effort to discover new anti-cancer drugs we investigate the efficacy of the product peroxides formed by the auto-oxidation of tetracycle 9H, 10H-indolizino [1, 2-b] indole-1one derivatives (2a-d) by a novel mechanism of azainolate/enamine intermediates as shown by our previous publication ¹ like allylic carbanion². Few peroxides are known to have anticancer activity due to the formation of peroxy radical ³. In addition, indoles are also known to have rich biological activity ⁴. With this known facts we have tested the anticancerous activity of the peroxides (7a - d, 10a – d, 12a – d) formed by the auto-oxidation of the synthesized indolizino-indole derivatives (2a -d) in the presence of base/Mel¹ or with only base and without methylating agent. The peroxides were isolated by silica gel column chromatography and tested against colon HT – 29, nasopharyngeal carcinoma HONE- 1 and BM -1 & breast carcinoma MX – 1 $^{5 \cdot 9}$ in vitro. The result is encouraging and many of the peroxides are found out to be inhibitor of cancerous cell growth in vitro. The IC₅₀ that is the concentration of drug needed to inhibit 50% cell growth is found out in low micromolar range. We anticipate that these peroxides

have a potency to be used as anti-cancerous agent. We are working on further study of these peroxides to find a clear cut picture of mode of action to use them as drug candidates.

MATERIAL AND METHODS

Compounds **2a** – **d** were synthesized by a known procedure using Fischer Indole ¹⁰ synthesis. Chemical required were purchased from Aldrich, Mecrk. Column chromatography was performed using silica gel G60 (70 – 230 mesh, Merck), thin layer chromatograph preparative layer chromatography were performed on silica gel GF₂₅₄ pre-coated aluminum foil (Merck). ¹HNMR was observed in Bruker 400 MHz instrument.

EXPERIMENTAL

The products were subjected to autooxidation procedure as given below.

Procedure for auto-oxidation without methylating agent

A solution of the indolizino indole (2a - d, 5 mmole) in dry DMF (50 mL) and dry potassium carbonate (2.05 g, 15 mmol) was heated at 80 ^o C for three h. The resulting solution was filtered and the solid was filtered out. From the filtrate DMF was distilled out and the product was extracted by ethyl acetate washing with water and dried by sodium

sulphate. The product was isolated by silica gel column chromatography eluting with 1 % methanol in ethyl acetate. The product is extremely unstable and the after repeated crystallization we got a very little pure product. Procedure for oxidation auto without methylating agent : A solution of the indolizino indole (2a - d, 5 mmole) in dry DMF (50 mL), dry potassium carbonate (2.05 g, 15 mmol) and methy iodide (15 mL, 24 mmol) was heated at 80 $^{\circ}$ C for three h. The resulting solution was filtered and the solid was filtered out. From the filtrate DMF was distilled out and the product was extracted by ethyl acetate washing with water and dried by sodium sulphate. Different products were isolated in different polarity of the eluting solvents. Compounds 9a - d were eluted first with hexane/EA (3:1, v/v) followed by 11a -d, 10a - d, 7a - d and 12a -d.

Biological assays and cytotoxic assay

The effect of the peroxides on cell growth were determined in all human tumor cells (i.e., colon HT – 29, nasopharyngeal carcinoma HONE- 1 and BM -1 &breast carcinoma MX – 1), in a 72 h. incubation, by XTT –tetrazolium assay, as prescribed in references ⁵⁻⁹. After the addition of phenazine methosulphate-XTT solution at 37 $^{\circ}$ C for 6 h. absorbance at 450 and 630 nm was detected on a microplate reader. 6 to 7 different concentrations of each compound were used. The IC₅₀ dose effect was calculated by a median relationship using a computer program.

¹H NMR and Purification Data 10-Hydroperoxy-2-cyano-3-methyl-1methoxy-indolizino[1,2-b]-indole (7a)

Eluted by Hexane : EA = 1 : 2 (v/v), ¹H NMR (DMSO d₆) δ 2.45 (3H, s), 3.86 (3H, s), 6.40 (1H, s), 7.21 (1H, t, *J* = 7.6 Hz), 7.37 (1H, t, *J* = 7.2 Hz), 7.54 (1H, d, *J* = 8.4 Hz), 7.65 (1 H, d, *J* = 8 Hz), 12.37 (1H, s, exch.), 12.48 (1H, s, exch.).

10-Hydroperoxy-2-cyano-3,7,8-trimethyl-1methoxy-indolizino[1,2-b]-indole (7b)

Eluted by Hexane : $\vec{EA} = 1 : 2 (v/v)$, ¹H NMR (DMSO d₆) δ 2.40 (3H, s), 2.44 (3H, s), 2.50 (3H, s), 4.02 (3H, s), 6.44 (1H, s), 7.05 (1H, brs), 7.35 (1H, brs),11.69 (1H, s, exch.), 12.38 (1H, s, exch.).

10-Hydroperoxy-2-cyano-3,6,8-trimethyl-1methoxy-indolizino[1,2-b]-indole (7c)

Eluted by Hexane : EA = 1 : 2 (v/v), ¹H NMR (DMSO d₆) δ 2.40 (3H, s), 2.45 (3H, s), 2.50 (3H, s), 4.02 (3H, s), 6.43 (1H, s), 7.09 (1H, s), 7.21 (1H, s), 7.54 (1H, d, *J* = 8.4 Hz),12.19 (1H, s, exch.), 12.42 (1H, s, exch.).

10-Hydroperoxy-8-fluoro-2-cyano-3-methyl-1-methoxy-indolizino[1,2-b]-indole (7d)

Eluted by Hexane : EA = 1 : 2 (v/v), ¹H NMR (DMSO d₆) δ 2.41 (3H, s), 4.22 (3H, s), 6.41 (1H, s), 7.26 (2H, m), 7.44 (1H, m), 13.22 (1H, s, exch.), 13.42 (1H, s, exch.).

9-*N*-methyl-10-hydroperoxy-2-cyano-3methyl-1-methoxy-indolizino[1,2-b]-indole (8a)

Eluted by Hexane : EA = 2 : 3 (v/v), ¹H NMR (DMSO d₆) δ 2.45 (3H, s), 3.86 (3H, s), 4.01 (3H, s), 6.40 (1H, s), 7.21 (1H, t, *J* = 7.6 Hz), 7.37 (1H, t, *J* = 7.2 Hz), 7.54 (1H, d, *J* = 8.4 Hz), 7.65 (1 H, d, *J* = 8 Hz), 12.47 (1H, s, exch.).

9-*N*-methyl-10-methylperoxy-2-cyano-3methyl-1-methoxy-indolizino[1,2-b]-indole (9a)

Eluted by Hexane : EA = 3 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.39 (3H, s), 3.18 (3H, s), 3.84 (3H, s), 4.19 (3H, s), 6.37 (1H, s), 7.24 (1H, d, J = 8.4 Hz), 7.48 (2H, d, J = 7.6 Hz), 7.74 (1H, d, J = 8.4 Hz).

9-*N*-methyl-10-methylperoxy-2-cyano-3,7dimethyl-1-methoxy-indolizino[1,2-b]-indole (9b)

Eluted by Hexane : EA = 3 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.35 (3H, s), 2.40 (3H, s), 2.44 (3H, s), 3.20 (3H, s), 3.39 (3H, s), 3.83 (3H, s), 6.43 (1H, s), 7.10 (1H, brs), 7.35 (1H, brs).

9-*N*-methyl-10-methylperoxy-2-cyano-3,7dimethyl-1-methoxy-indolizino[1,2-b]-indole (9c)

Eluted by Hexane : EA = 2 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.35 (3H, s), 2.45 (3H, s), 2.56 (3H, s), 2.83 (3H, s), 3.22 (3H, s), 4.34 (3H, s), 6.40 (1H, s), 7.05 (1H, s), 7.21 (1H, s).

9-*N*-methyl-10-methylperoxy-8-fluoro-2cyano-3-methyl-1-methoxy-indolizino[1,2b]-indole (9d)

Eluted by Hexane : EA = 2 : 1(v/v), ¹H NMR (DMSO d₆) δ 2.41 (3H, s), 2.83 (3H, s), 3.68 (3H, s), 4.22 (3H, s), 6.41 (1H, s), 7.26 (2H, m), 7.44 (1H, m).

9-*N*-methyl-10-hydroperoxy-2-cyano-3methyl-1-methoxy-indolizino[1,2-b]-indole (10a)

Eluted by Hexane : EA = 2 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.45 (3H, s), 4.07 (3H, s), 4.14 (3H, s), 6.42 (1H, s), 7.27 (1H, d, *J* = 8.4Hz), 7.65 (2H, m), 7.72 (1H, d, *J* = 8.2 Hz), 12.42 (1H, s, exch.).

9-*N*-methyl-10-hydroperoxy-2-cyano-3,7dimethyl-1-methoxy-indolizino[1,2-b]-indole (10b)

Eluted by Hexane : EA = 1 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.35 (3H, s), 2.50 (3H, s), 3.82 (3H, s), 4.00 (3H, s), 6.94 (1H, s), 7.33 (1H, d, J = 8 Hz), 7.58 (2H, d, J = 8 Hz), 11.72 (1H, s, exch.).

9-*N*-methyl-10-hydroperoxy-2-cyano-3,6dimethyl-1-methoxy-indolizino[1,2-b]-indole (10c)

Eluted by Hexane : EA = 2 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.37 (3H, s), 2.41 (3h, s), 2.56 (3H, s), 3.79 (3H, s), 3.96 (3H, s), 6.46 (1H, s), 7.07 (1H, s), 7.21 (1H, s), 12.26 (1H, s, exch.).

9-*N*-methyl-10-hydroperoxy-8-fluoro-2cyano-3-methyl-1-methoxy-indolizino[1,2b]-indole (10d)

Eluted by Hexane : EA = 2 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.43 (3H, s), 3.78 (3H, s), 4.22 (3H, s), 6.43 (1H, s), 7.27 (2H, m), 7.46 (1H, m), 13.29 (1H, s, exch.).

10-Hydroperoxy-2-cyano-1-hydroxyindolizino[1,2-b]-indole (11a)

Eluted by Hexane : EA = 1 : 1 (v/v), ¹H NMR (DMSO d₆) δ 2.50 (3H, s), 3.82 (3H, s), 3.93 (3H, s), 3.96 (3H, s), 7.27 (1H, m), 7.36 (1H, s), 7.41 (1H, m), 7.69 (1H, d, *J* = 8.4 Hz), 7.98 (1H, d, *J* = 8.8 Hz).

10-Hydroperoxy-2-cyano-3-methyl-1hydroxy-indolizino[1,2-b]-indole(12a)

Eluted by 1 % MeOH in EA (v/v), ¹H NMR (DMSO d₆) δ 2.45 (3H, s), 3.86 (3H, s), 6.40 (1H, s), 7.21 (1H, t, *J* = 7.6 Hz), 7.37 (1H, t, *J* = 7.2 Hz), 7.54 (1H, d, *J* = 8.4 Hz), 7.65 (1 H, d, *J* = 8 Hz), 12.37 (1H, s, exch.), 12.48 (1H, s, exch.), 13.01 (1H, s, exch.).

10-Hydroperoxy-2-cyano-3,7,8-trimethyl-1hydrooxy-indolizino[1,2-b]-indole (12b)

Eluted by 1 % MeOH in EA (v/v), ¹H NMR (DMSO d₆) δ 2.40 (3H, s), 2.44 (3H, s), 2.50 (3H, s), 4.02 (3H, s), 6.44 (1H, s), 7.05 (1H, brs), 7.35 (1H, brs), 11.69 (1H, s, exch.), 12.38 (1H, s, exch.), 13.72 (1H, s, exch.).

10-Hydroperoxy-2-cyano-3,6,8-trimethyl-1hydrooxy-indolizino[1,2-b]-indole (12c)

Eluted by 1 % MeOH in EA (v/v), ¹H NMR (DMSO d₆) δ 2.40 (3H, s), 2.45 (3H, s), 2.50 (3H, s), 4.02 (3H, s), 6.43 (1H, s), 7.09 (1H, s), 7.21 (1H, s), 7.54 (1H, d, J = 8.4 Hz),12.19 (1H, s, exch.), 12.42 (1H, s, exch.), 13.23 (1H, s, exch.).

10-Hydroperoxy-8-fluoro-2-cyano-3-methyl-1-hydroxy-indolizino[1,2-b]-indole (12d)

Eluted by 1 % MeOH in EA (v/v), ¹H NMR (DMSO d₆) δ 2.41 (3H, s), 4.22 (3H, s), 6.41 (1H, s), 7.26 (2H, m), 7.44 (1H, m), 13.22 (1H, s, exch.), 13.42 (1H, s, exch.), 13.41 (1H, s, exch.).

RESULT AND DISCUSSION

The tetracyclic 9H, 10H-indolizino [1, 2-b] indole-1-one (2a - d) and its derivatives are known¹ to be auto-oxidized in the presence of base and alkylation agent to produce a series of compounds (7a - d, 8a - d, 9a - d, 10a - d, 11a - d, 12a - d, Scheme 1). Among these compounds the peroxides (7a - d, 10a - d, **12a - d**) are expected to be active towards propagation of anti-cancerous cells³ due to chance of formation of peroxy radical. But the yield of peroxide is very poor may be due to the formation of methyl peroxides in the presence of methylating agent. With the aim of synthesizing enough amount of peroxide (12a - d) for testing it against different cancerous cell lines in vitro, we carried out the same reaction without methylating agent and with only base (Scheme 2). The yield of 12a **d** improved a little amount. The list of compounds synthesized was given in tabular form in Table 1 and the yields are given in Table 2 (when the reaction is carried out with methylating agent) and Table 3 (when the reaction is carried out without methylating agent and with base only).

The results clearly shows that yield of the peroxides (**12a - d**) improve a little bit when the reaction was carried out without methylating agent. In addition we have little amount of methylated peroxides (**7a–d**, **10a–d**) in our hand. The isolation and purification of the products are very complicated. After repeated purification by silica gel column chromatography and preparative thin layer chromatography we got enough purity of the compound to test the biological activity.

With all the required peroxides in our hand we have tested *in vitro* anti-cancerous activity assay against cancerous cell lines of colon HT – 29, nasopharyngeal carcinoma HONE- 1 and BM -1 & breast carcinoma MX – 1 by published procedure ⁵⁻⁹. The result is shown in Table 4.

From the result (Table 4) it is apparent that among these peroxides compounds 7a - dand compounds 12a - d are most potent against almost all the cancerous cell lines in the range of nanomolar IC₅₀ values. It is obvious that these two series of peroxides are having unprotected indole NH group, as shown from the structure of these products in Scheme 1 and Table 1 and Table 2. This may be attributed due to better binding of these compounds with the cancerous cell lines. This is our speculation and to prove it we need to carry out further study. The peroxides **10a–d** are showing potent anti-cancerous activity against HT - 29 and MX - 1 cell lines, but almost inactive against HONE - 1 and BM – 1 though.

We also tried to find the *in vivo* activity of these compounds against nude mice grafted with cancerous cell. However, due to the formation of very small amount of compound and the extreme instability and also insolubility of these peroxides we could not do it. We also attempted to test *in vivo* biological activity on nude mice grafted with cancerous cell with the indolizino-indole derivatves 2a - d, but these compounds are also extremely insoluble in solvent like DMSO. So a conclusive data we could not find.

The cause of anti-cancer activity of the peroxide *in vitro* may be due to cleaving of DNA in cancerous cell by the eroxyradical ³ as radicals are known DNA cleaving agents $\frac{12}{12}$. The peroxides may be damaging DNA of different cell lines by binding with it and thus blocking cell propagation in cell cycle. However, this conclusion we arrived by literature survey of published research of previous researchers ¹². To get a conclusive report, a more detailed study with more stable peroxides is needed.

With all these studies it is confirmed though, that the peroxides formed by the autooxidation of indolizino-indole derivatives (2a - d) have a potential to be treated as an anticancerous agent as shown from the *in vitro* study of these compounds against different cancerous cell lines. A detailed study to find a complete SAR is needed. Also *in vivo* anti cancerous activity of these candidates are also needed. Still we can conclude that most of the peroxides are potent against different cancerous cell line *in vitro*.

CONCLUSION

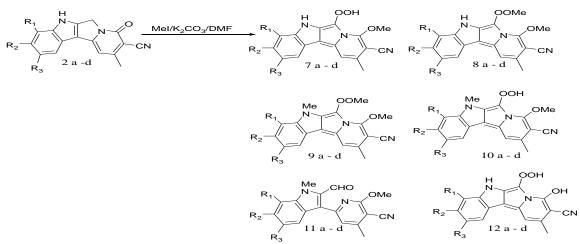
From the above data we may conclude that many of the peroxides formed by the autooxidation of tetracyclic 9H, 10H-indolizino [1, 2-b] indole-1-one (2a - d) have a potential to be used as an anti-cancerous agents as shown by their *in vitro* anti-cancerous activity assay against different cancerous cell lines. The *in vitro* activity assay was done against different cancerous cell lines. Among those peroxides few derivatives of 10-Hydroperoxy-2-cyano-3-methyl-1-hydroxy-indolizino[1,2-b]-

indole (12a - d) are found out to be most active with IC₅₀ in nanomolar range. It is also observed that presence of indole NH and peroxo OH is useful to increase the potency of these derivatives.

It is also found out that the yield of peroxide (12a - d) of the auto-oxidation of tetracyclic 9H, 10H-indolizino [1, 2-b] indole-1-one (2a - d) can be improved if the reaction is carried out without methylating agent, though isolation of pure product is really tough and needs a rigorous purification.

ACKNOWLEDGEMENT

The author is grateful to UGC for funding the project {No. F. PSW-009/07-08 (ERO)} and No. F. PSW-036/15-16 (ERO). The author is also highly indebted to Prof. Tsann Long Su, Institute of Biomedical Sciences, Academia Sinica, Taipei, Taiwan, for allowing him to carry out the initial research work at Academia Sinica as Post-Doctoral Fellow. My work with Dr. Su opens up a route to study with these peroxides.



Scheme. 1: Auto-oxidation using methylating agents



Scheme. 2: Auto-oxidation without methylating agent

Table 1: List of compound synthesized				
Number of Compound	R ₁	R ₂	R₃	
2a, 7a, 8a, 9a, 10a, 11a, 12a	Н	Н	Н	
2b, 7b, 8b, 9b, 10b, 11b, 12b	Me	Me	Н	
2c, 7c, 8c, 9c, 10c, 11c, 12c	Me	Н	Me	
2d, 7d, 8d, 9d, 10d, 11d, 12d	F	H	Н	

Table 2: Yield of different compound produced				
by auto-oxidation with Mel/K ₂ CO ₃				

by auto-oxidation with wei/R ₂ CO ₃				
Compound	Product	Yield %	Yield of 7a after rigorous purification %	
2a	7a	52		
	8a	1		
	9a	5	12	
	10a	8] 12	
	11a	11		
	12a	1		
	7b	60		
	8b	0 (ND)		
2b	9b	12	10	
	10b	8	10	
	11b	0 (ND)		
	12b	0 (ND)		
	7c	55		
	8c	0 (ND)		
2c	9c	15	8	
20	10c	11	8	
	11c	0 (ND)		
	12c	0 (ND)		
	7d	58		
	8d	0 (ND)		
2d	9d	14	5	
	10d	12	5	
	11d	0 (ND)		
	12d	0 (ND)	1	

Table 3: Yield of different compound produced by auto-oxidation with base $K_2 \dot{C} O_3$

Starting material	Product	Isolated Yield (%)
2a	12a	16
2b	12b	20
2c	12c	17
2d	12d	15

Compound	Inhibition of cancerous cell growth (IC ₅₀ µM)*			
Number	HT - 29	HONE - 1	BM - 1	MX -1
7a	0.098	0.36	0.45	0.067
7b	0.085	0.86	1.1	0.092
7c	0.067	0.097	0.88	0.12
7d	0.081	1.2	0.96	0.079
10a	1.2	>100	45	1.8
10b	2.8	>100	34	0.11
10c	1.7	87	68	0.32
10d	0.97	>100	>100	0.17
12a*	0.079	0.21	0.23	0.051
12b*	0.087	0.34	0.11	0.068
12c*	0.073	0.55	0.31	0.081
12d*	0.056	0.78	0.56	0.091

Table 4: Inhibition of different cell lines (IC₅₀ μ M)

* Purity of these compounds is a matter of

concern and result may vary due to this.

REFERENCES

- Bhattacharya G, Su TL, Chen KT and Chia CM. Synthesis and autooxidation of new tetracyclic 9H, 10 H,indolizino[1,2-b]indole-1-ones. J Org Chem. 2001;66:426-432.
- 2. Barton DHR and Jones DL. Autooxidation in basic media, Part IV, hydrocarbon autooxidation. J Chem Soc. 1965;3563–3570.
- Doroshow JH and Davies KJ. Redox cycling of anthracyclines by cardiac mitochondria. II. Formation of superoxide anion, hydrogen peroxide, and hydroxyl radical. J Biol Chem. 1986; 20:3008-3014.
- Brown RK. The chemistry of heterocyclic compounds. Houlihan. J.H. Edited, Interscience: New York. 1972.
- Chou TC. The Median-Effect Principle and the CombinationIndex for Quantitation of Synergism and Antagonism. Synergism and Antagonism in ChemotherapyChou. T.C.; Rideout. D. Academic Press, New York. 1991.
- 6. Scudiero DA, Shoemaker RH, Paull KD, Monks A, Tierney S, Nofziger TH, Currens MJ, Seniff D and Boyd MR. Evaluation of soluble

tetrazolium/formazan assay for cell growth and drug sensitivity in culture using human and other tumor cell lines. Canc Res. 1988;48:4827-4833.

- Rastogi K, Chan JY, Pan WY, Chen CH, Chou TC, Chen LT, Su TL. Antitumor AHMA linked to DNA minor groove binding agents : synthesis and biological evolution. J Med Chem. 2002; 45:4585 – 4593.
- Chou TC and Talalay P. Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. Adv Enz Reg. 1984;22:27-55.
- 9. Chou TC. The median-effect principle and the combination index for quantitation of synergism and antagonism. In synergism and antagonism in ahemotherapy. 1991.
- 10. Fischer E and Jourdan F. Ueber die hydrazine der brenztraubensäure, Berichte. 1883;16: 2241–2245.
- 11. Sinha BK. Free radicals in anticancer Drug Pharmacology. Chem Biol Int. 1989;69:293–317.
- 12. http://en.wikipedia.org/Freer adical damage to DNA.