

MANGO LEAF UTILIZED AS A LOW-COST ADSORBENT FOR THE REMOVAL OF POTASSIUM ION FROM AQUEOUS SOLUTION

Chaudhary B*, Desai V and Shah M

Pioneer Pharmacy Degree College, Near Ajwa Cross Road, N.H.-8, Ajwa Nimeta Road, At & Post Sayajipura, Vadodara – 390019, Gujarat, India.

ABSTRACT

The potential of mango leaves (*Mangifera indica*), as a low-cost adsorbent for the removal of K⁺ ions from aqueous solution was investigated in this study. The influences of pH, contact time, initial metal concentration, agitation rate, particle size and effect of biosorbent dosage were studied in batch experiments at room temperature. Maximum sorption for metal was found to be at pH 6.5. Process parameters which include initial metal ion concentration, adsorbent dosages, initial pH and temperature of solution were varied in order to evaluate their influence on the adsorption process. The results obtained indicate that the adsorption of K⁺ on MLP is better at higher metal ion concentrations and lower adsorbent dosages. Langmuir and Freundlich isotherm models were fitted to the biosorption of K⁺ ions on the mango leaves, it giving correlation coefficient of 0.9991. Langmuir model fitted the equilibrium data better, giving correlation coefficient of 0.9999 and a maximum adsorption capacity of 42.5 mg/g. This indicates monolayer coverage on adsorbent. The results showed that mango leaves have the potential to be applied as alternative low-cost biosorbent in the remedy of metal contamination in waste water.

Keywords: Mango leaf powder, K⁺ ions, biosorption, adsorption, waste water.

INTRODUCTION

In moving toward the new era of science and technology, the world has seen major transformations in numerous aspects of life. At the heart of this, tremendous advancement lies the rapid industrialization occurring at various parts of the globe which causes detrimental effects for mankind and animals. One widespread phenomenon which has drawn much attention is the contamination of toxic metals such as copper, lead, zinc, nickel, potassium & chromium in the aquatic environment, which are sourced from chemical industries such as petrochemicals, refineries, fertilizers, pulp and paper¹.

³. Heavy metal contamination may cause serious health problems such as cancer and brain damage, due to the accumulation in living tissues and organs².

⁴. Generally, metals could be divided into four distinguished categories which are toxic metals, strategic metals, precious metals and radionuclides. Among these

toxic metals are the ones associated for causing serious environmental threats, making its removal from the aquatic environment essential. From the various types of toxic metals present in wastewater, Potassium was chosen for this biosorption studies with regard to its wide use in industry and potential pollution impact. In industrial waste, Potassium mainly appears to be in the form of the monovalent K(I), which is more toxic than the metal itself. This is because it is soluble in water in its ionic form and can easily absorb into living organisms. High doses of Potassium in the aquatic environment generate toxicological concerns as it can deposit into the brain, liver, pancreas and myocardium³. Thus, K⁺ concentration of wastewater should be reduced to a value of at least 0.2 to 8 mg/L as per WHO guideline for drinking water. The increase of metal bearing effluents into the aquatic environment has caused progressive developments in wastewater treatment. Atypical wastewater treatment plant is divided into

everal areas and the removal of metallic pollutants is performed mainly in the tertiary stage. Current developed methods in this stage include filtration, ion exchange, membrane separation, nutrient stripping and adsorption^{1,7}. However, these methods have some limitations due to technical and economical constraints. The ion exchange process needs a high operational cost because it involves the use of excessive amount of reagent for its resin regeneration. It also requires a high capital cost for the equipment used. Meanwhile, the membrane process application is limited by the condition of the membrane used. This membrane tends to be unstable in salty or acidic conditions and fouled by organic or inorganic substances present in the wastewater. The major disadvantage for the precipitation process is that it generates a toxic sludge that requires careful disposal as per regulations. For electrolysis, its inefficiency at low concentrations which causes incomplete metal ion recovery which is one of its disadvantages⁸. Due to these factors, research has been conducted to study the use of natural adsorbents as an alternative, based on the economical and environmental point of view. The most current technique employed for metal removal from wastewater is biological adsorption or biosorption. "Biosorption" is the term given to the passive sorption and/or complexation of metals by biomass⁹. It is a reversible process where the concentration of metal ions in solution decreases due to adsorption onto the solid phase until a dynamic equilibrium between the aqueous and solid phase is established. This technology which utilizes natural biomass materials is very effective for the detoxification of metal-bearing industrial effluents. The biosorption process involves several mechanisms that differ qualitatively and quantitatively, depending on the origin of the biomass, the species used and its processing¹⁰. These mechanisms are generally based on physico-chemical interactions between metal ions and functional groups present on the cell surface, which include ion exchange, complexation, electrostatic attraction and microprecipitation¹¹. The process of biosorption has many attractive features including the selective removal of metals over a broad range of pH and temperature, its rapid kinetics of adsorption and desorption and low capital and operation cost. Due to higher affinity of the sorbent for the sorbate species, the later is attracted and removed by different mechanisms. The process continues till equilibrium is established between amount of solid-bound sorbate species and its portion remaining in solution. Various types of biomass have been used as the biosorbent for the removal of toxic metals. Among these, plant leaves are chosen in this study as they are proposed to be natural, simple and cheap biosorbents for the efficient removal of several heavy metal ions. They can be easily found and are a renewable source.

Plant leaves are also non-toxic and biodegradable. After the biosorption process, they are expected to precipitate and become sediments which can be disposed safely. Mulch plant leaves were used for the biosorption of cadmium, lead and copper ions. The results proved that the performance of the leaves is close to the efficiency of using activated carbon. The results also showed that oven-dried leaves have better performance compared to the naturally dried ones. The functional group which is commonly found in plant leaves is carboxylate⁹. Available literature involving the use of mango leaf powder (MLP) for the biosorption of heavy metals is quite limited. Particularly, no study involving MLP for the removal of Potassium ion was found. One paper was found in which the researcher used several test plant materials including MLP as the biosorbent for the removal of lead ions¹²⁻¹⁸.

AIM AND OBJECTIVES

1. To find an inexpensive and effective adsorbent to replace commercial methods in removing heavy metals from aqueous effluent.
2. To investigate the possible use of mango leaves as a low cost adsorbent for removal of K^+ ions.
3. To study the effect of pH, effect of initial concentration of K^+ ions, contact time, effect of agitation rate, effect of particle size and effect of chelating agent on the adsorption of K^+ ions on mango leaves.

MATERIAL AND METHODS

Preparation of biosorbent

Mature and fresh mango leaves as shown in figure 1 were collected from local trees and washed thoroughly by using distilled water to clean them from dirt and impurities. After that, the leaves were sun-dried for a day on a perforated tray until the leaves turned brownish in colour. The leaves were then dried further in an oven (Labtech, EIE-101) for 24 hours until the leaves became crisp. After drying, the leaves were ground by a mechanical grinder (Premir, Xprees 750), to a constant size of 80 μ m and the resulting MLP was kept in a glass bottle ready for further experiments.

Preparation of adsorbate

The potassium stock solution (1000 mg/L) was prepared, which was purchased from Suvidhinath laboratories. Necessary dilutions were done to obtain 50 mg/L, 75 mg/L and 100 mg/L of potassium ion solution. Standard concentrations of 10, 20, 30 mg/L of K^+ solutions were also prepared for calibration purposes. The initial and final metal concentrations were an

alyzed via Flame Photometry (Chemiline, Cat#: CL410) using the flame at specific condition. The pH of the K^+ solution was adjusted by adding 0.1M HCl or 0.1M NaOH and the measurement was done by pH meter (Chemline, ATCCL120).

Biosorption experiments

The biosorption of K^+ from an aqueous solution was investigated in batch biosorption experiments. The MLP with different dosages (0.4, 0.6, 0.8, 1.0 and 1.2 g/L) were added each in 5 conical flasks containing 100 mL of 100 mg/L KCl solution. 5 mL of the KCl solution was withdrawn and added in volumetric flask that contained 45 mL of deionised water for the purpose of dilution.

After that, the batch shake flask experiments were performed using a sonicator (Toshcon ultrasonic cleaner, Toshniwal Instrument Pvt. Ltd. Ajmer) with 75 rpm at 30°C for 3 hours. For the first hour, the samples were withdrawn every 15 minutes. For the following hours, the same procedure was repeated at every 30 minutes interval. The samples were retested for their K^+ ion concentration with the Flame Photometry.

For pH experiments, the pH of the solution was adjusted to 2.5, 4.5, 8.5 and 10.5 by using 0.1M HCl and 0.1M NaOH. Five different sets of experiments with variable concentration of K^+ solution, pH and temperature were performed.

The amount of material adsorbed per unit mass of adsorbent (mg/g) is given in equation (1):

$$q = \frac{C_0 - C_t}{m} \dots \dots \dots (1)$$

Where, C_0 is the initial concentration of adsorbate (mg/L) and C_t is the concentration of adsorbate at time t (mg/L).

The adsorption efficiency can be expressed as percentage adsorption of metal ion per equation (2):

$$\% \text{ adsorption} = \frac{C_0 - C_t}{C_0} \times 100\% \dots \dots \dots (2)$$

Where, C_0 and C_t are the initial adsorbate concentration (mg/L) and the adsorbate concentration at time t (min) respectively.

Equilibrium studies

Equilibrium data, commonly known as adsorption isotherms, are basic requirements for the design of adsorption systems. Classical adsorption models, Langmuir (1918) and Freundlich (1907), were used to describe the equilibrium between potassium ions on the MLP at constant temperature.

The Langmuir equation is valid for a monolayer sorption on a homogeneous surface with a finite number of identical sites and when there are no interactions between

the sorbed species. The linear form of Langmuir equation is given in equation (3):

$$\frac{C_e}{q_e} = \frac{1}{q_m} * C_e + \frac{1}{Kl} (qm) \dots \dots \dots (3)$$

where C_e (mg/L) is the equilibrium concentration of adsorbate, q_e (mg/g) is the quantity of adsorbed material (mg/g) at equilibrium, K_l is the Langmuir equilibrium constant related to the energy of sorption ($L \cdot mg^{-1}$) and q_m is the maximum amount of metal ions per unit weight of MLP to form a complete monolayer on the surface bound at high C_e . It also represents a practical limiting adsorption capacity when the surface is fully covered with the metal ions and assists in the comparison of adsorption performance, particularly in cases where the sorbent did not reach its full saturation in experiments¹⁶⁻¹⁸.

The empirical Freundlich equation applied to multilayer sorption on a heterogeneous surface and can only be employed in the low to intermediate concentration ranges. The Freundlich equation is given in equation (4):

$$\log q_e = \frac{1}{n} \log C_e + \log K_f \dots \dots \dots (4)$$

where the K_f ($mg \cdot g^{-1}$) and n (value between 0 and 1) are the Freundlich constant characteristic of the system. K_f and n are indicators for adsorption capacity and adsorption intensity, respectively^{17,19-26}.

RESULTS AND DISCUSSION

1) Effect of pH:

The pH of the adsorbate solution is considered one of the most important factors affecting the biosorption process. This factor is capable of influencing not only the binding site dissociation state, but also the solution chemistry of the target metal in terms of hydrolysis, complexation by organic and/or inorganic ligands and redox potentials.

The range of pH tested in this experiment was from 2.5 to 10.5. Solutions with pH lower than 2.5 was also not suitable as for these solutions, the surface active sites of the adsorbent would be protonated, resulting in a competition of K^+ and H^+ ions for the same surface active sites which would result in a low potassium uptake^{4,27-29}.

In the present study, the pH was adjusted in the range of 2.5-6.5 by using dilute H_2SO_4 and 8-10.5 by using NaOH. The experiments were carried out at 50 mg/L, 75 mg/L, 100 mg/L of initial metal ion concentration with 1.2 g/L MLP dosage. Draw the graph of pH vs % adsorption according to table 1. From Fig. 2, it can be observed that the adsorption is highest for pH

6.5 followed by pH. These results were also present in many previous studies which involved a maximum adsorption from pH 4.5 to 5.5. This trend was expected as at low pH values, most of the binding sites on the cell surface would have positive charges making the binding and desorption of the metal ions unattractive to the sites³⁰⁻³².

The competition of K^+ with H^+ ions for appropriate sites on the adsorbent surface lowers the adsorption of K^+ ions. However, as the pH of the solution increases, this competition weakens, where K^+ ions replace H^+ ions bound to the adsorbent since increasing number of protons are dissociated from functional groups on the cell wall. In addition, at higher pH, there will be a change in the dominant functional groups responsible for binding on the cell wall, with carboxylate groups and imine groups being the most important ones for pH < 5 and pH 6 respectively³⁵. More negative groups were then made available on the adsorbent surface as pH increases.

The negatively charged adsorbent surface increases the electrostatic attraction between positively charged adsorbate and negatively charged adsorbent particles and therefore, leading to an increase in the adsorption of K^+ ions. However, further increase in the pH will cause the metal to form precipitates.

2) Effect of Time:

The effect of contact time was studied at different initial metal ion concentrations and MLP dosages. It can be clearly observed that the percentage of adsorption generally increased until time reached 120 minutes. After this time, there was a drop in the adsorption percentage. Thus, 120 minutes was chosen as the optimum time where the adsorption reached equilibrium.

The highest percentage of adsorption was found to be 31.86%, 27.36% and 54.54% for concentration of 50 mg/L, 75 mg/L and 100 mg/L respectively. From the concentration experiments, 100 mg/L of K^+ was found to be optimal for the dosage of MLP used. The relationship of adsorption efficiency with time at varied MLP dosages is depicted in Fig. 3 according to table 2. The results reveal a similar trend, where MLP removal is higher at the beginning (for the first 45 minutes) for all the dosages used.

This is due to the larger surface area of MLP available during the initial contact time. As the surface adsorption sites become exhausted, the uptake rate is controlled by the rate at which the adsorbate is transported from the exterior to the interior sites of the adsorbent particles. Similar results were reported using wheat shell as the biosorbent. Figures show that the initial metal ion concentration and MLP dosage did not affect the time for the adsorption to reach its optimum time of 120 minutes. After the optimum time, both plots also show a decrease in the removal efficiency. This indicates

that the desorption process may have occurred, as adsorption is a reversible process³⁶⁻⁴⁵.

3) Effect of Metal Ion Concentration

The initial metal ion concentration plays an important role towards the performance of a batch biosorption. The effect can be studied from the results of experiments at constant dosages of MLP. As shown in Fig. 4, the metal uptake of different dosages of MLP increased as the initial metal ion concentration increased from 50 to 100 mg/L, where the maximum value of metal uptake was observed to be from 42.5 mg/g to 7.5 mg/g respectively. These values were obtained at the optimum contact time of 120 minutes and MLP dosage of 0.4 g/L. The value of 42.5 mg/g is the maximum metal uptake, or the maximum adsorption capacity that was achieved by the biosorption of K^+ on MLP in this study. This observation was also in line with the research performed by two other researchers⁴⁶⁻⁴⁸.

The result is expected as the initial metal ion concentration functions as the driving force to overcome mass transfer resistances between the aqueous and solid phases. In addition, the increase in the initial metal ion concentration also increased the number of collisions between the metal ion and the biosorbent, hence increasing the metal uptake. Thus, it could be concluded that the highest metal uptake would occur at the highest metal concentration, which is 100 mg/L, at optimal conditions⁴⁹.

4) Effect of Biosorbent Dosage

The influence of the biosorbent dosage on the process was examined by using five different dosages at 0.4, 0.6, 0.8, 1.0 and 1.2 g/L. Fig. 5 shows the relationship of metal uptake with the biosorbent dosage at K^+ ion concentration of 100 mg/L and time of 120 minutes according to table 4. It can be observed that the increase in the biosorbent dosage causes a decrease in the metal uptake. Similar results were also observed for K^+ concentrations of 50 mg/L and 75 mg/L. This phenomenon is due to the splitting effect of flux (concentration gradient) between the adsorbate and biosorbent with increasing biomass concentration causing a decrease in amount of metal ion adsorbed per gram of biomass.

Another factor is that at high sorbent dosages, the available metal ions are insufficient to cover all the exchangeable sites on the biosorbent, usually resulting in a low metal uptake⁵⁰. Besides, it might be due to the formation of aggregation during biosorption causing a decrease in the effective adsorption area when the biomass concentration increases. Thus, it could be concluded that for this study, the highest metal uptake occurs at the lowest biosorbent dosage (0.4 g/L) if other process conditions are kept constant⁵¹.

5) Adsorption isotherms

Many models have been proposed to explain adsorption equilibria, but the most important factor is to have applicability over the entire range of process conditions. The most widely used isotherms for solid-liquid adsorption are the Langmuir and Freundlich isotherms. Both of these isotherms relate the adsorption density, q_e (metal uptake per unit weight of adsorbent) to equilibrium adsorbate concentration in the bulk fluid phase. Langmuir isotherm model is valid for monolayer adsorption onto a surface containing a finite number of identical sites. This isotherm is derived from the assumption that a maximum adsorption corresponds to a saturated monolayer of solute molecules on the adsorbent surface, with constant energy of adsorption, and no transmigration of adsorbate in the plane of the surface⁵². The correlation coefficient that was obtained from the Langmuir plot as shown in Fig. 6 is 0.999. This trend is also observed for adsorption at different MLP dosages and initial metal ion concentration. These values prove that the adsorption is favorable for a Langmuir isotherm. Freundlich isotherm gives the relationship between the equilibrium liquid and solid phase capacity based on multilayer adsorption (heterogeneous surface). The Freundlich isotherm is based on the assumption that the adsorption sites are distributed exponentially with respect to the heat. Based on Fig. 7, the correlation coefficient that was found from the Freundl

ich plot is 0.9991. This high value indicates that the adsorption is favorable for a Freundlich isotherm. In addition, the adsorption intensity, n , which was found to be 0.625, further proves this finding. This is as this value is smaller than 1. Meanwhile, the Freundlich constant, K_F was found to be 1.628. These results also indicate that the MLP surface is heterogeneous in the long range, but may have short range of uniformity⁵⁴.

CONCLUSION

The following conclusions can be drawn based on the investigation of K^+ adsorption by MLP:

1. The maximum adsorption capacity of K^+ on MLP was 206.85 mg/g which was obtained with 0.4 g/L MLP dosage, 100 mg/L K^+ concentration and pH of 4.5.
2. The optimum contact time for the maximum adsorption capacity of K^+ on MLP was 120 minutes.
3. The metal uptake of K^+ on MLP decreased with increasing MLP dosage.
4. The metal uptake and adsorption efficiency of K^+ on MLP increased with increasing metal ion concentration.
5. The experimental data for the adsorption of K^+ on MLP fits well for the Freundlich isotherm.

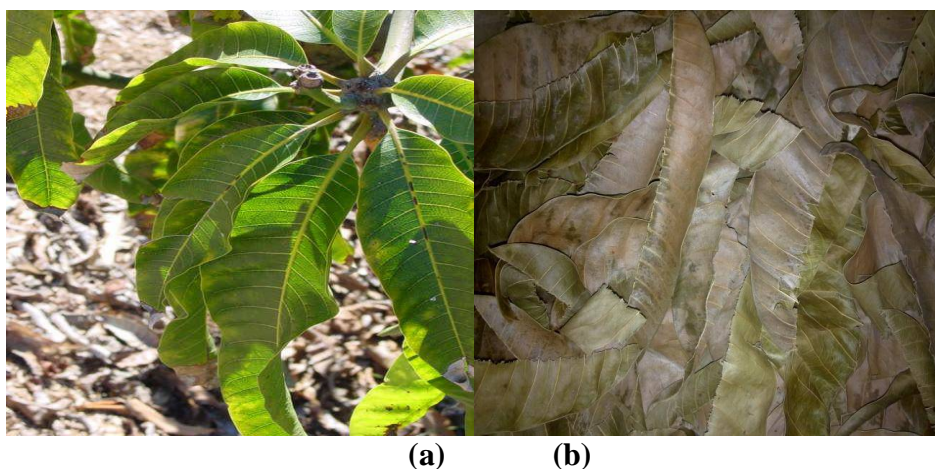
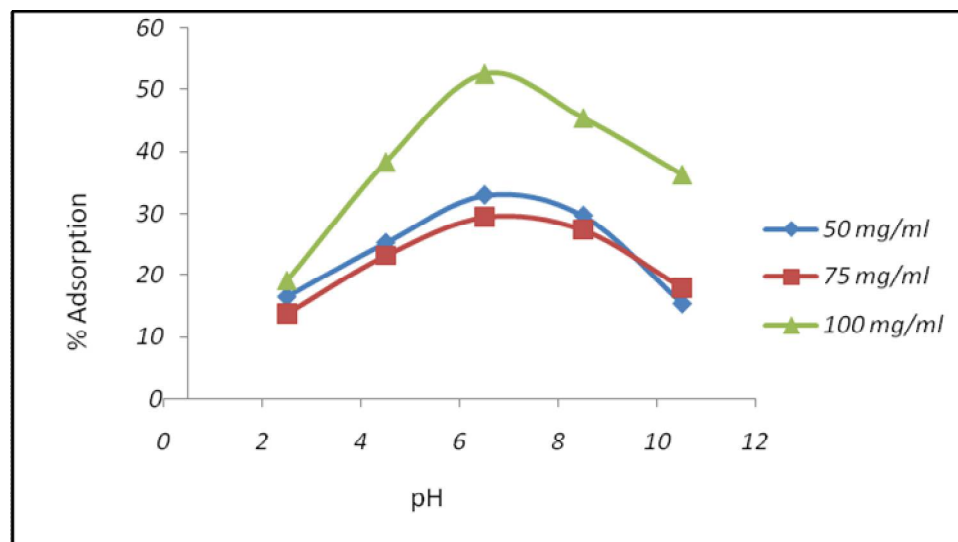


Fig. 1: (a) Natural mango leaves (b) Dried mango leaves

Table 1: Effect of pH in K⁺ ion concentration

Sr.no	pH	Conc. Of k ⁺ ion (mg/ml)		
		50	75	100
1	2.5	16.48	13.68	19.19
2	4.5	25.27	23.15	38.38
3	6.5	32.96	29.47	52.52
4	8.5	29.67	27.36	45.45
5	10.5	15.38	17.89	36.36

Fig. 2: Plots of adsorption of K⁺ ion(50, 75, 100 mg/L) on MLP (1.2 g/L) using various pHTable 2: Effect of time in K⁺ ion concentration

S.N o.	Time (min)	Conc. of K ⁺ ion (mg/ml)		
		50	75	100
1	0	91	95	99
2	60	12.08	9.47	8.08
3	120	31.86	27.36	54.54
4	180	24.17	20	17.17

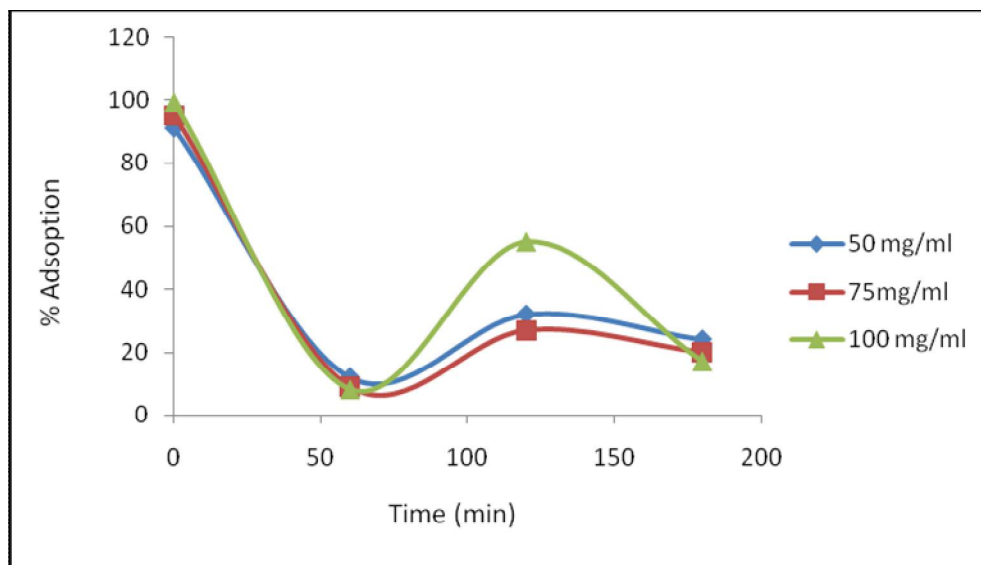


Fig. 3: Plots of adsorption of K⁺ ion(50, 75, 100 mg/L) on MLP (1.2 g/L)using various time interval (60, 120 and 180 mins)

Table 3: Effect of concentration in metal ion uptake

Concentration	Metal uptake				
	I	II	III	IV	V
50	5.1	6	8.12	19.2	37.9
75	6	4	10.6	20.21	40.6
100	7.5	7	12.5	23.33	42.5

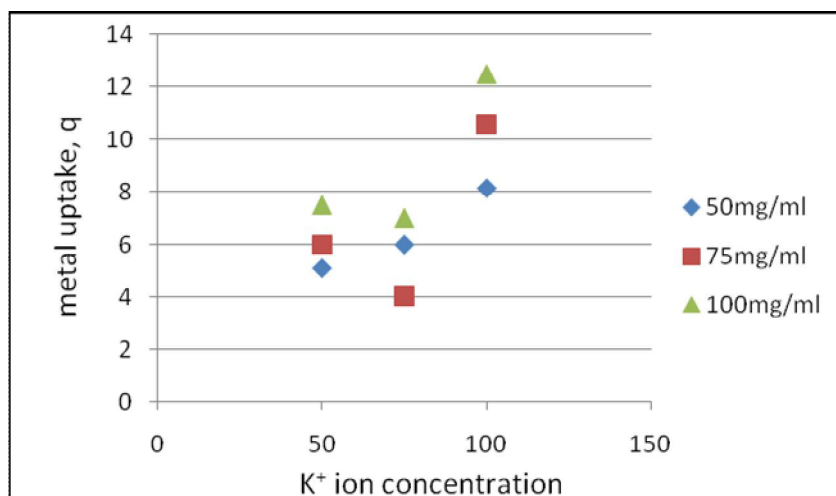
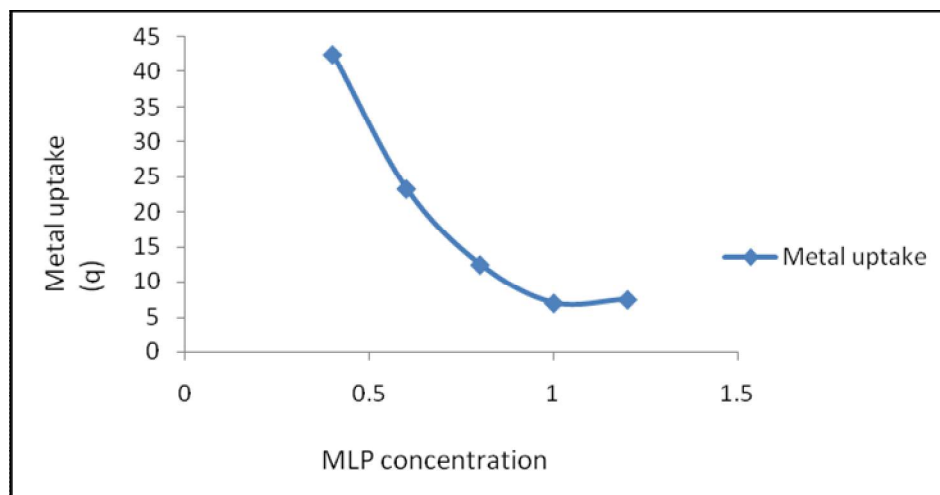


Fig.4: Plots of metal uptake, q against K⁺ ion concentration with different MLP dosages at 30°C

Table 4: Effect of biosorbent dosage form in metal ion uptake

S.No.	Con. Of MLP (mg/ml)	Metal Uptake(q)in %
1	0.4	42.5
2	0.6	23.23
3	0.8	12.5
4	1.0	7
5	1.2	7.5

**Fig.5: Plot of metal uptake, q against MLP dosage at 100 mg/L K⁺ concentration at 30°C****Table 5: Langmuir adsorption isotherm**

Ce/Cq (mg/ml)	Ce (mg/ml)
1.36	42.5
0.91	23.33
0.65	12.5
0.52	7
0.54	7.5

Table 6: Freundlich adsorption isotherms

log qe	log Ce
2.64	1.62
2.47	1.36
2.3	1.09
2.16	0.84
2.17	0.87

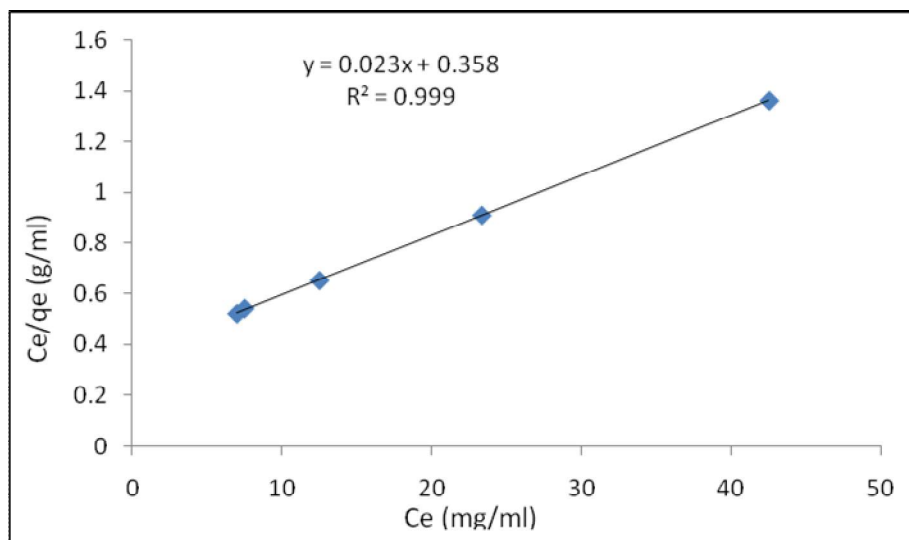


Fig. 6: Langmuir plot for adsorption of K^+ on MLP with dosage of 0.4 g/L and at 30°C

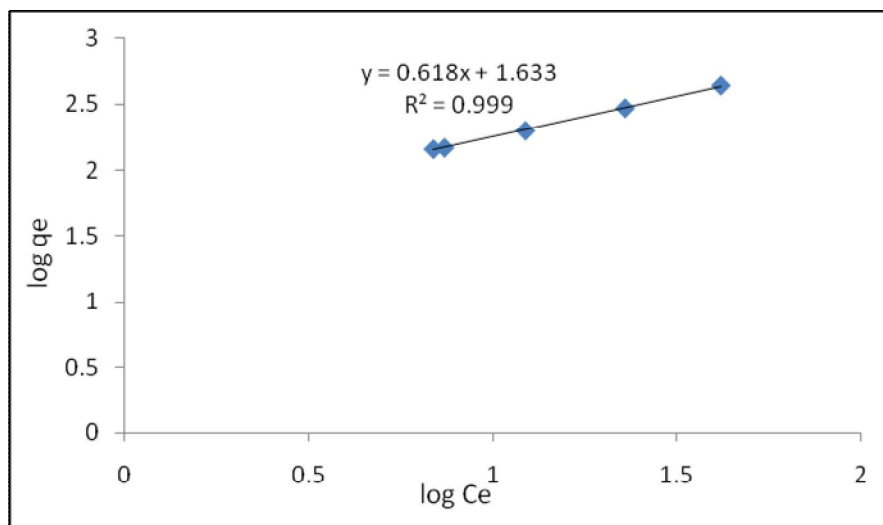


Fig. 7: Freundlich plot for adsorption of K^+ on MLP with dosage 0.4 g/L and at 30°C

REFERENCES

- Sethu, Vasanthi; Goye, K.S.; Iffah, F.R.; Khoo, C.M.; Andresen, John M. Adsorption characteristics of Cu(II) ions in aqueous solution using *Mangifera indica*. 2010; 5(2): 262-278.
- Ong Pic Sheen. Utilization of Mango leaf as low-cost adsorbent for the removal of Cu(II) ion from aqueous solution. 2011.
- K.S. Rao, M. Mohapatra, S. Anand, P.Venkateswarlu. Review on Cadmium removal from aqueous solutions. 2010; 2(7): 81-103.
- Kaewsarn P. and Yu Q. Cadmium (II) removal from aqueous solutions by pretreated biomass of marine alga *Padinasp.*, Environ. Pollut. 2001; 112(1): 209–213.
- Mukhopadhyay M., Role of surface properties during the biosorption of copper by pretreated *Aspergillus niger* biomass, Colloids and Surfaces, Physicochem. Eng. Asp. 2008; 329(2): 95-99.
- Davis T. A., Volesky B. and Vieira, R.H.S.F. Sargassum seaweed as biosorbent for heavy metals, Water Res. 2000; 34(1): 4270-4278.

7. Lu D., Cao Q., Li X., Cao X., Luo F. and Shao W. Kinetics and equilibrium of Cu(II) adsorption onto chemically modified orange peel cellulose biosorbents, *Hydro. metallurgy*.2008; doi:10.1016/j.hydro.2008;
8. Dang V. B. H., Doan H. D., Dang-Vu T. and Lohi A. Equilibrium and kinetics of biosorption of cadmium (II) and copper (II) ions by wheat straw, *Biores. Tech.*2008; 100(2): 211-219.
9. Mack C., Wilhelmi B., Duncan J. R. and Burgess J. E. Biosorption of precious metals, *Biotech. Adv.*2008; 25(1): 264-271.
10. Volesky B. and Holan Z. R. Biosorption of heavy metals. *Biotech. Prog.*1990;1(2): 235-250.
11. Schiewer S. and Volesky B. Modeling the Proton-Metal Ion Exchange in Biosorption. *Environ. Sc. Tech.*1995; 29(2): 3049-3058.
12. Shaban W., Al Rmalli, Abdella A. Dahmani, Mohamed M. Abuein and Amar A. Gleza. Biosorption of mercury from aqueous solutions by powdered leaves of castor tree. *J. Ha. Mate.*2007; 152(2): 955-959.
13. Sarma J., Sarma A. and Bhattacharyya K. G. Biosorption of commercial dyes on Azadirachtaindicaleaf powder: A case study with a basic dye Rhodamine B, *Ind. Eng. Chem. Res.*2008; 47(2): 5433-5440.
14. Pamukoglu Y. M. and Kargi F. Effects of operating parameters on kinetics of copper (II) ion biosorption onto pretreated waste sludge (PWS). *Enzy. Micr. Tech.* 2007; 42(2): 76-82.
15. Cochrane E. L., Lua S., Gibb S. W. and Villaescusa I. A comparison of low-cost biosorbents and commercial sorbents for the removal of copper from aqueous media, *J. Ha. Mate. B.*2006; 137(2): 198-206.
16. Sheng P. X., Ting Y. P., Chen J. P. and Hong L. Sorption of lead, copper, cadmium, zinc, and nickel by marine algal biomass: characterization of biosorptive capacity and investigation of mechanisms, *J. Colloid Interf. Sc.*2004;275(3): 131–141.
17. Yu Q., Matheickal J. T., Yin P. and Kaewsarn P. Heavy metal uptake capacities of common marine macroalgal biomass, *Water Res.*1999; 33(6): 1534–1537.
18. Hawari A. H. and Mulligan C. N. Biosorption of lead(II), cadmium(II), copper(II) and nickel(II) by anaerobic granular biomass, *Biores. Tech.*2006; 97(1):692 – 700.
19. Equilibrium, kinetics and thermodynamic parameters of the batch biosorption of nickel(II) ions onto *Chlorella vulgaris*, *Proc. Biochem.*2002; 38(1): 89-99.
20. Ajmal M., Rao R.A.K., Ahmad R. and Ahmad J., Adsorption studies on Citrus reticulata (fruit peel of orange): removal and recovery of Ni(II) from electroplating wastewater, *J. Haz. Mate.B.*2000; 79:117-1131.
21. Bulut Y. and Tez, Z., Adsorption studies on ground shells of hazelnut and almond, *J. Ha. Mate.*2007; 149(1): 35-41.
22. Qin F., Wen B., Shan X. Q., Xie Y. N., Liu T., Zhang S. Z. and Khan S. U., Mechanisms of competitive adsorption of Pb, Cu (pH = 4), and Cd on peat, *Env.Poll.* 2006; 144(1): 669–680.
23. Nasernejada B., Zadehb T. E., Poura B. B., Bygia M. E. and Zamani A. Comparison for biosorption modeling of Cr, Cu, Zn wastewater by carrot residues, *Proc. Biochem.*2005; 40(2): 1319 –1322.
24. Wong K. K., Lee C. K., Low K. S. and Haron M. J., Removal of Cu and Pb by tartaric acid modified rice husk from aqueous solutions, *Chemos.*2003; 50(1): 23–28.
25. Saeed A., Iqbal M. and Akhtar M. W. Removal and recovery of lead(II) from single and multimetal (Cd, Cu, Ni, Zn) solutions by crop milling waste (black gram husk), *J. Ha. Mate. B.* 2005; 117(2): 65–73.
26. Sun G. and Shi W. Sunflower stalks as adsorbents for the removal of metal ions from wastewater, *Ind. Eng. Chem. Res.*1998; 37(2): 1324–1328.
27. Reddad Z., Gerente C., Andres Y. and LeCloirec P. Adsorption of several metal ions onto a lowcost

- adsorbent, kinetic and equilibrium studies, *Environ. Sc.Tech.* 2002; 36(1): 2067-2073.
28. Keskinan O., Goksu M. Z. L., Yuceer A., Basibuyuk M. and Forster, C.F. Heavy metal adsorption characteristics of a submerged aquatic plant *Myriophyllum spicatum*, *Proc. Biochem.* 2003; 39(2): 179-183.
29. Ozsoy H. D. and Kumbur H. Adsorption of Cu(II) ions on cotton ball, *J. Ha.Mate.* 2006; 136(1): 911-916.
30. Bhatti H.N., Khalid, R. and Hanif, M.A. Dynamic biosorption of Zn(II) and Cu(II) using pretreated *Rosa gruss* and *teplitz* (red rose) distillation sludge, *Chem. Eng. J.* 2008; 148(1): 434-443.
31. Ngeontae W., Aeungmaitrepirom, W., Tuntulani, T. and Imyim, A. Highly selective preconcentration of Cu(II) from seawater and water samples using amidoamidoxime silica, *Talanta.* 2008; 43(2): 1004-1010.
32. Demirbas E., Dizge N., Sulak M. T. and Kobya M. Adsorption kinetics and equilibrium of copper from aqueous solutions using hazelnut shell activated carbon, *Chem. Eng. J.* 2008; 148(1): 480-487.
33. Jiang Y., Pang H., and Liao B. Removal of copper(II) ions from aqueous solution by modified bagasse, *J. Ha. Mate.* 2007; 148(1): 1-9.
34. Pavan F. A., Lima I. S., Lima E. C, Airoldi, C. and Gushikem Y. Use of Ponkan mandarin peels as biosorbent for toxic metals uptake from aqueous solutions, *J. Ha. Mate. B.* 2006; 137(2): 527-533.
35. Al-Subu M. M., Salim R., Abu-Shqair I. and Swaileh K. M. Removal of dissolved copper from polluted water using plant leaves: Effects of acidity and plant species, *Revi. Interna. de Contam.Ambi.* 2001; 17(1): 91-96.
36. Wang G. Biosorption of heavy metal ions from aqueous solutions by nonliving water hyacinth roots, Ph.D. Dissertation, University of Nevada, Reno, UMI Company, 1995.
37. Bulut Y. and Tez, Z. Adsorption studies on ground shells of hazelnut and almond, *J. Ha. Mate.* 2007; 149(1): 35-41.
38. K. L. Dorris, Yu B., Zhang, Y., Shukla, A. and Shukla, S. S. The removal of heavy metal from aqueous solutions by sawdust adsorption- removal of copper, *J. Ha. Mate., B.* 2000; 80(2): 33-42.
39. Basci N., Kocadagistan E. and Kocadagistan B. Biosorption of copper (II) from aqueous solutions by wheat shell *Desalination.* 2004; 164: 135-140.
40. Ekmekyapar F., Ali A., Kemal Y. B. and Avni C., Biosorption of copper (II) by non living lichen biomass of *Cladonia rangiformis*, *J. Ha. Mate. B.* 2006; 137(1): 293-298.
41. Ozsoy H. D. and Kumbur H. Adsorption of Cu(II) ions on cotton ball, *J. Ha. Mate.* 2006; 136(1): 911-916.
42. Al-Asheh S. and Duvnjak Z. Adsorption of copper by canola meal, *J. Ha. Mate.* 1996 ;48 : 83-93.
43. Al-Asheh S. and Banat F. Adsorption of copper and zinc by oil shale, *Environ. Geo.* 2001; 40(6): 693-698.
44. Al-Asheh S. and Duvnjak Z. Sorption of cadmium and other heavy metals by pine bark, *J. Hazard. Mate.* 1997; 56(2): 35 -51.
45. Ulmanu M., Marañón E., Fernández Y., Castrillón L., Anger I. and Dumitriu, D. Removal of copper and cadmium ions from diluted aqueous solutions by low cost and waste material adsorbents, *Wat. Air Soil Pollu.* 2003; 142(2): 357- 373.
46. Tsui M. T. K., Cheung K. C., Tam N. F. Y. and Wong M. H., A comparative study on metal sorption by brown seaweed, *Chemospher.* 2006; 65(2): 51-57.
47. King P., Anuradha, K., Beena Lahari S., Prasanna Kumar, Y., and Prasad V.S.R.K. Biosorption of zinc from aqueous solution using *Azadirachta indica* bark: Equilibrium and kinetic studies, *J. Ha. Mate.* 2008; 152(2): 324-329.
48. Gupta V. K., Srivastava S. K. and Mohan D. Equilibrium uptake, sorption dynamics, process optimization and column operations

- for the removal and recovery of Malachite Green from wastewater using activated carbon and activated slag, *Ind. Eng. Chem. Res.* 1997; 36(1): 2207-2212.
49. Sawalha M. F., Peralta-Videa J. R. and Romero-Gonzalez, J. Thermodynamic and isotherm studies of the biosorption of Cu(II), Pb(II), and Zn(II) by leaves of saltbush (*Atriplex canescens*), *J. Chem. Thermody.* 2007; 39(1): 488-492.
50. Vijayaraghavan K., Palanivelu K. and Velan, M. Biosorption of copper (II) and cobalt(II) from aqueous solutions by crab shell particles, *Biores. Tec.* 2006; 97(2): 1411-1419.
51. Ekmekyapar F., Ali A., Kemal Y. B. and Avni C. Biosorption of copper (II) by non living lichen biomass of *Cladoniarangiformis hoffm*, *J. Ha. Mate. B.* 2006; 137(1): 293-298.
52. Kalavathy M. H., Karthikeyan T., Rajgopal S. and Miranda L. R. Kinetic and isotherm studies of Cu(II) adsorption onto H₃PO₄-activated rubber wood sawdust, *J. Coll. Interf. Sc.* 2005; 292(1): 354–362.
53. Gaballah I. and Kilbertus G. Recovery of heavy metal ions through decontamination of synthetic solutions and industrial effluents using modified barks, *J. Geochem. Explor.* 1998; 62(1): 241-286.
54. Dada, A.o, Olalekan; A.P, Olatunya, A.M, DADA, O. Langmuir, Freundlich, Temkin and Dubini-Radushkevich Isotherms Studies of Equilibrium Sorption of Zn²⁺ into Phosphoric acid Modified Rice Husk. 2012; 3(1) :38-45.