

PREPARATION AND EVALUATION OF SUSTAINED RELEASE MICROBEADS OF NORFLOXACIN USING SODIUM ALGINATE

Anuranjita Kundu*

Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, West Bengal University of Technology, Kolkata, India.

ABSTRACT

Oral controlled drug delivery systems represent the most popular form of sustained drug delivery systems for the obvious advantages of oral route of drug administration. Such systems release the drug with constant or variable release rates. The oral controlled release systems shows a typical pattern of drug release in which the drug concentration is maintained in the therapeutic window for a prolonged period of time (sustained release), thereby ensuring sustained therapeutic action. They are used as single dosage form. Present work involves preparation and evaluation of sustained release of microspheres of norloxacin employing sodium alginate as natural polymer. The technique employed for microencapsulation of the drug is ionotropic gelation.

Keywords: Microbeads, Ionotropic Gelation, Natural Polymers, Sustained Release.

INTRODUCTION

The goal of any ideal drug delivery system is to provide a therapeutic amount of drug to the proper site in the body to achieve prompt response, and thus maintain the desired drug concentration. Such a conceptualized ideal drug delivery can be possible with intravenous infusion of drug at the site of action over a desired period of time. However, such a drug therapy requires hospitalization of the patient and needs to be administered by a qualified person. Therefore, among the routes preferred for the administration of dosage forms, the oral route presents a series of attractive advantages for the administration of the drug compounds, including the avoidance of pain and discomfort associated with injections and elimination of possible infections caused by the use of needles. More over, oral dosage forms are less expensive to produce, because they do not need to be manufactured under sterile conditions. In recent years, scientific and technological advancements have been made in the research and development of rate

controlled oral drug delivery systems. These formulations are designed to deliver the drugs at a controlled and predetermined rate, thus maintaining their therapeutically effective concentration in systemic circulation for prolonged periods of time. A great majority of controlled release formulations have been formulated in the form of tablets. However, wide physiological and environmental variations in the gastrointestinal tract with respect to the surface area of absorption, pH, fluidity, rate of transit time, presence of food constituents and co-administered drugs may influence gastric emptying. If a controlled release product is formulated in the form of tablet which keeps its integrity throughout the gastrointestinal tract, then the location of the tablet will vary under different circumstances. This will lead to the variation in the rate of drug delivery to the systemic circulation. Compared with single unit dosage form, multiunit controlled release drug delivery systems like microcapsules and microspheres are

becoming popular as they pass through the gut as if a solution avoiding the vagaries of gastric emptying and different transit rates¹ spread over a large area of absorbing mucosa preventing exposure to high drug concentration² and release drug in a more predictable manner³.

Controlled release dosage forms cover a wide range of prolonged action formulations, which provide continuous release of their active ingredients at a predetermined rate and for a predetermined time. The majority of these formulations are designed for oral administration. The most important objective for the development of these systems is to furnish an extended duration of action and thus assure greater patient compliance.

MATERIALS AND METHODS

Norfloxacin was obtained as a gift sample from Dey's Medical Stores (mfg.) LTD., Sodium Alginate was supplied by Loba chemie pvt. Ltd., Calcium chloride (CAS 10035-04-8) was purchased from National chemicals.

PREPARATION OF NORFLOXACIN LOADED MICROBEADS^{4,5,6}

The microparticles were prepared by ionotropic gelation technique. At first, sodium alginate (1%, 2%, 3%w/v) was dispersed uniformly in 10 ml of warmed distilled water using mechanical stirring maintaining speed at fixed rpm so that the aqueous mucilage of sodium-alginate was obtained.^{7,8} This aqueous mucilage was then kept for 5-10 mins. To remove any air bubbles that may have been formed during stirring process. Then to this aqueous mucilage of sodium alginate, the accurately weighed drug (norfloxacin) was added very slowly and stirred the whole system at fixed rpm for about 5 minutes and as a result, the drug was uniformly dispersed in the aqueous mucilage of the sodium-alginate. The sodium-alginate drug dispersion was added drop-wise via a needle fitted with a 10ml syringe into 100ml of 4% calcium chloride solution.⁹ After incubating for a predetermined time, the gelled beads were separated by filtration and washed with distilled water. Then the microparticles were dried at the room temperature. The following experimental parameters were varied:

Table 1: Composition of Norfloxacin Microparticles

FORMULATION CODE	DRUG LOADING (%W/V)	SODIUM ALGINATE (% W/V)	CALCIUM CHLORIDE (% W/V)
A1	30 %	1%	4%
A2	30 %	2 %	4 %
A3	30 %	3 %	4 %

Three formulations of microparticles (A1,A2 & A3) were prepared in identical manner only changing the amount of sodium alginate in these as respectively 1% w/v, 2% w/v & 3% w/v. These three formulations were subsequently subjected to evaluation tests.

EVALUATION STUDIES

DRUG ENTRAPMENT EFFICIENCY:^{10,11}

Accurately weighed amount of norfloxacin loaded A1, A2, A3 beads were kept in 100ml of USP phosphate buffer solution of pH 6.8 and kept for 24 hours. The solution was filtered and an aliquot following suitable dilution was assayed spectrophotometrically (UV-VIS Spectrophotometer, Thermo Spectronic UV-1) for norfloxacin at 270 nm. The drug entrapment efficiency was determined using the following relationship:

$$\text{Drug Entrapment Efficiency (DEE)} = \frac{\text{experimental drug content}}{\text{theoretical drug content}} * 100$$

Table 2: Drug Entrapment Efficiency of prepared formulations

S.No.	Formulation Code	% Drug Entrapment Efficiency
1	A1	83.78
2	A2	88.56
3	A3	92.13

SCANNING ELECTRON MICROSCOPY (SEM)

The surface morphology of microparticles can be analyzed by scanning electron microscopy (SEM). The shape, surface morphology of the alginate microparticles can be investigated using SEM (HITACHI E-1010). The sample was deposited on brass hold and sputtered with gold by using (model no.) fine coat ion sputter device. The SEM photographs were taken with the scanning electron microscope at required magnification at room temperature. The working distance of 39 mm was maintained and acceleration voltage used was 15 kv.

PARTICLE SIZE DETERMINATION

Particle size of the alginate beads was determined using an optical microscope using a compound microscope (OLYMPUS 01C). A standard stage micrometer was used to calibrate the optical micrometer.

Table 3: Effect of alginate concentrations on particle size

S.No.	Formulation Code	Diameter (μm) (% \pm S.D. n=3)
1	A1	523.85 (\pm 46.742)
2	A2	525.3833 (\pm 46.929)
3	A3	542.6 (\pm 47.125)

SWELLING STUDY

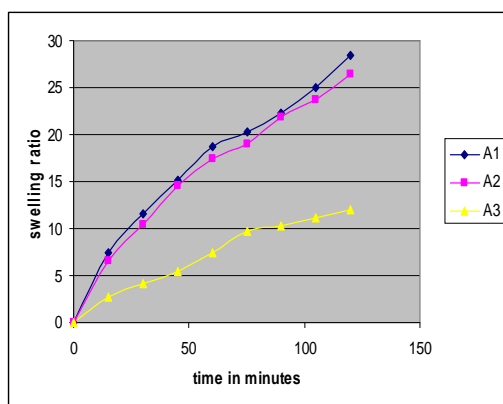
The swelling characteristics of blank ALG and drug loaded ALG beads were determined by immersing about 10mg (weigh in Precisa XB 600M-C) of dried test samples to swell in 50 ml of a solution at pH 1.2 and pH 6.8 medium. At specific time intervals, samples were removed from the swelling medium and were blotted with a piece of tissue paper to absorb excess water on surface. The swelling ratios (Q) of test samples were calculated from the following expression:

$$Q = (W_s - W_d) / W_d$$

Where W_s is the weight of the swollen test sample and W_d is the weight of the dried test sample.

Table 4: Swelling characteristics of different formulations

Time in min.	A1	A2	A3
0	0	0	0
15	7.47	6.56	2.78
30	11.57	10.46	4.16
45	15.15	14.64	5.48
60	18.68	17.37	7.49
75	20.34	19.05	9.68
90	22.32	21.89	10.27
105	24.98	23.75	11.12
120	28.46	26.49	11.97

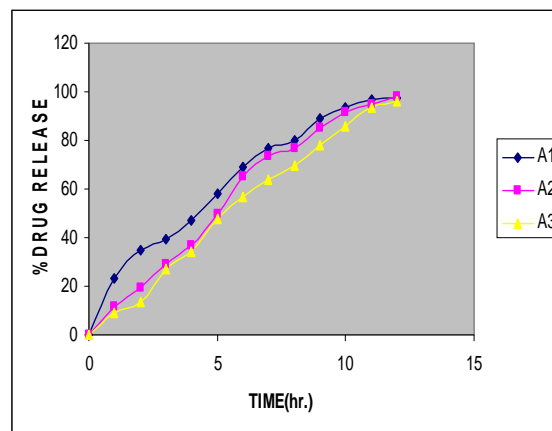
**Graph 1: Swelling study for A1, A2, A3 formulations****IN-VITRO DRUG RELEASE STUDY^{12,13}**

The in vitro dissolution study was carried out using six station dissolution rate apparatus USP at 50 rpm AT $37 \pm 0.5^\circ \text{C}$. The dissolution

medium consists of simulated intestinal fluid for 5 hours. 5ml of the fluid containing the micropellets in the apparatus were removed every 30 minutes and an equivalent amount of fresh dissolution equilibrated at the same temperature was replaced. The sample withdrawn was diluted, filtered and analyzed at 278 spectrophotometrically. The plot of percentage cumulative drug release against time (hrs.) is given as under.

Table 5: In-Vitro Drug Release Studies for Prepared Microparticles

Time in hours	A1 (Cumulative % Release)	A2 (Cumulative % Release)	A3 (Cumulative % Release)
0	0	0	0
1	23	11.45	9.03
2	34.54	19.56	13.56
3	39.56	28.98	27.12
4	46.96	36.78	34.37
5	58.32	49.94	47.68
6	69.07	65.47	56.76
7	76.63	73.56	63.79
8	79.98	76.78	69.99
9	88.75	85.37	78.36
10	93.59	91.35	85.89
11	96.88	94.58	93.56
12	97.83	97.34	96.19

**Graph 2: In-vitro drug release study for A1, A2, A3 formulations****RESULT AND DISCUSSION**

Microbeads of Norfloxacin were prepared by ionotropic gelation technique and different evaluation parameters were assessed, with a view to obtain oral control release of Norfloxacin. In the drug entrapment efficiency study, decrease in initial alginate concentration decreased DEE at a given curing time (10

min.) and CaCl_2 concentration (4%). Decrease in initial alginate concentration provides lesser number of binding sites of alginate for Ca^{2+} ions resulting in the formulation of a less compact gel membrane which, in turn, increases influx of Ca^{2+} ions leading to decrease in DEE. The particle size of the prepared different batches of microparticles were presented in Table No. 3. The microparticles were found to be in the size range of 500-550 μm . It was observed that in these prepared Norfloxacin microbeads, with the increase in alginate percentage the distribution of particle size shifts to the higher value due to increase in the initial viscosity of the medium. In order to find the effect of cross linking on the release rates of Norfloxacin from the matrix, swelling was studied in terms of percentage of water uptake by the beads. However the swelling property of the beads was measured in terms of percentage of water uptake by the beads at a particular time interval. The results of the percent of water uptake by different beads are presented in GRAPH No.1. It was observed that from the GRAPH NO. 1, the higher the amount of sodium alginate in the beads, the lower the swelling rate. The in-vitro drug release studies of different formulations cumulative percentage drug release was observed in the range of 96.19-97.83. The in-vitro drug release profile was mentioned in the Table No. 5. The formulations A1, A2, A3 containing 1%, 2%, 3% sodium alginate respectively showed a release of 97.83%, 97.34% and 96.19% after 12 hours. This shows that more sustained release was observed with the increase in percentage of sodium alginate. The best formulation was observed as A3, by the observation of all results of the three formulations Norfloxacin microbeads.

CONCLUSION

This study reveals that oral controlled release of Norfloxacin can be successfully achieved by ionotropic gelation technique using sodium alginate as polymer. Prepared microbeads shown higher drug entrapment efficiency and prolonged release characteristics. Norfloxacin release from microbeads was influenced by different alginate concentrations. Among the different formulations of microbeads, A3 was estimated as best formulation because this formulation drug release was observed that drug was released in controlled manner.

ACKNOWLEDGEMENTS

The author is thankful to Dey's Medical Stores (Manufacturing) Limited, Kolkata for providing the gift sample of Norfloxacin.

REFERENCES

- 1) Beckett AH, Alternative routes of drug administration and new drug delivery systems: Towards better safety of drug and pharmaceutical products. North Holland: Elsevier North Holland Biomedical Press, 1980;247-263.
- 2) Devis SS, Hardy JG, Taylor MJ, Whalley DR and Wilson CG. Comparative study of gastrointestinal transit of a pellet and tablet formulation. *Int J Pharm.* 1984;21:167-177.
- 3) Follonier N and Doelkar E., Biopharmaceutical comparison of oral multiple-unit and single-unit sustained release dosage forms. *STP Pharmaceutical Sciences.* 1992; 2:141-158.
- 4) Lym-Ly and Wan-LS. Propranolol Binding in Calcium Alginate Beads. *Drug Develop Indi. Pharm.* 1997; 23(10):973-980.
- 5) Manna A, Ghosh I, Goswami N, Ghosh LK and Gupta BK. Design and evaluation of an oral controlled Release Microparticulate Drug Delivery system of Nimesulide by Ionotropic Gelation Technique and Statistical Optimization by Factorial Analysis. *J Sci Ind Res.* 1999; 58(9):717-722.
- 6) Patil VB and Varsha B. Preparation and Evaluation of Sustained Release Nimesulide Microspheres prepared from sodium alginate. *Indian J Pharm Sci.* 2001;63(1):15-19.
- 7) Chowdary KPR and Srinivasa rao Y. Preparation and Evaluation of Mucoadhesive Microparticles of Indomethacin. *Indian J Pharma Sci.* 2003;65(1):49-52.
- 8) Nikhil O, Dhoot and Margaret A. 'Microencapsulated Liposomes in Controlled Drug Delivery: Strategies to Modulate Drug Release and Eliminate the Burst Effect. *J Pharm Sci.* 2003;92(3): 679-689.
- 9) Chowdary KPR and Srinivasa rao Y. Mucoadhesive Microparticles of Glipizide; Characterization, In-vitro and In-vivo Evaluation. *Indian J Pharm Sci.* 2003;65(3):279-284.
- 10) Srineevasa Rao B and Ramana murthy KV. Preparation and evaluation of flurviprofen Microcapsules by Emulsification Solvent Evaporation Technique'. *Indian Drugs,* 1996;33(8); 397-400.

- 11) Verma PRP, Neha Sharam and Lata Jha. Release Profile of Flurviprofen from Ointment Bases Through Cellulose Acetate Film. *The Indian Pharmacist*. 2003; 5: 57-59.
- 12) Murali Mohan Babu GV, Prasad CHDS and Himasankar K. Development of New Controlled Release Formulation of Flurviprofen; In vitro-In vivo correlation. *Indian J Pharm Sci*. 2002; 64(1): 37-43.
- 13) Udupa N and Setharaju G. Spectrophotometric Method of Analysis of Flurviprofen in Tablets, Plasma and Urine samples. *Indian Drugs*. 1989; 26(10):585-587.