

PHYTOCHEMICAL SCREENING AND ANTIBACTERIAL ACTIVITY OF ETHANOLIC EXTRACT OF TERMINALIA CATAPPAFLOWERS

T. Ramachandramoorthy^{1*}, M. Soji George¹, S. Balasubramaniyan²,
K. Rajasekar², L. Palanivelan², R. Govindharaju² and B. Jayalakshmi³

¹PG & Research Department of Chemistry, Bishop Heber College (Autonomous),
Tiruchirappalli, Tamil Nadu, India.

²Department of Chemistry, Government Arts College, Ariyalur, Tamil Nadu, India.

³Syed Ammal Engineering College, Ramanathapuram, Tamil Nadu, India.

ABSTRACT

TerminaliaCatappais a well known medicinal plant belonging to the family of *Combretaceae*. Due to its effective biological activities, it is used as a folk medicine in Southeast Asia. It is well known for Indian system of medicines. *TerminaliaCatappais* one of the plants having a rich Phytoconstituents like *Flavonoids, Sterols, Taninns and Saponins*.

The aim of the present study is to Prepare the ethanolic extract of the flowers of *TerminaliaCatappa*, to Identify the compounds present in the extract and to Evaluate the antibacterial activity of ethanolic extract. GCMS analysis helps to identify the compounds present in the extract. The antibacterial activity was done by well diffusion method. It shows good activity against *Staphylococcus aureus, Escherichia coli* and *Pseudomonas putida*.

Keywords: TerminaliaCatappa, Ethanolic extract, GCMS, Antibacterial activity.

INTRODUCTION

TerminaliaCatappais a well known traditional medicinal plant from the family of *Combretaceae*. It is one of the plants which are widely used in Ayurvedic medicinal systems. Various biological applications like anticancer, antioxidant, anti HIV, reverse transcriptase, antidiabetic, anti-inflammatory and hepatoprotective activity have been reported for various parts of this plant¹. The methanolic extract from the leaves of this plant shows very good antitumor activity against Ehrlich Ascetic Lymphoma [EAL] cell lines². The presence of various bioactive constituents like alkaloids, flavonoids, cardiac glycosides, tannins, saponins and cyanogenic glycosides in the decoction of *Terminaliacatappa* in place of water shows very good biochemical results which proves

the efficacy of the plant in medicinal systems³. Antimicrobial activity of the aqueous extract of leaves of *TerminaliaCatappa* was evaluated against *Klebsiellapneumoniae, Staphylococcus aureus, Escherichia coli* and *Candida albicans*. It shows good activity against *Klebsiellapneumoniae* and further it has more microbes potency than standard antibiotics like penicillin and ampicillin also reported⁴. Five carotenoids viz., violaxanthin, lutein epoxide, lutein, two lutein isomers and beta cryptoxanthin were isolated and characterized from the leaves of this plant⁵. The water extract of this plant leaves has better *in vitro* bacterial activity against the bacteria isolated from the aquatic animals like *Pasteurellapneumotopica* (0.8 mg/ml), *Photobacterium damsela* and *Enterococcus faecal*⁶. Two new flavones glycosides

apigenin 6-C-(2''-O-galloyl)-beta-D-glucopyranoside and apigenin 8-C-(2''-O-galloyl)-beta-D-glucopyranoside together with four known flavone glycosides called isovitexin, vitexin, isoorientin, and rutin were also isolated from the dried fallen leaves of *Terminalia catappa*⁷. Antioxidant and anthelmintic potential was evaluated from the leaves of *Terminalia catappa*⁸ and then the leaves extracts also inhibit MMP-9 expression and HCC cell metastasis⁹. Alcoholic and aqueous extract from the fruits of this plant shows very good antihyperglycemic activity in alloxan induced diabetic in rats¹⁰. From the seeds of this plant, essential oil was extracted and physicochemical characterization of this oil was evaluated¹¹. To be the part of research works on *Terminalia catappa*, this study shows the antibacterial activity and screening of compounds in the ethanolic extract of flowers of *Terminalia catappa* using well diffusion method and GC-MS.

MATERIALS AND METHODS

Collection and preparation of flower extract

Fresh flowers of *Terminalia catappa* were collected from Virudhunagar, Tamil Nadu, India during August 2015. About 100 ml of ethanol was added to about 50 grams of coarsely powdered flowers and the extraction carried out for 5 hours at 50°C in Soxhlet apparatus. The crude extract was pooled to the volume of 200 ml and then filtered using Whatman filter paper. Then the extract was concentrated in a rotator evaporator and kept in air tight amber color bottle. The non-soluble portion of the extracted solid remains in the thimble, and is discarded¹².

Phytochemical screening

The following preliminary phytochemical tests were carried out using the above mentioned ethanolic extract to detect the presence of different phytoconstituents. The phytochemical screenings were performed using standard procedures.

- a) **Test for terpenoids [Salkowski test]:** To small amount of the extract solution, 1 ml of chloroform and 5 ml concentrated H₂SO₄ was carefully added to form a layer. A reddish brown colouration of the interface indicates the presence of terpenoids¹³.
- b) **Test for flavonoids:** About 1 ml conc. HCl and piece of magnesium ribbon were added to 2 ml of the ethanolic extract. The development of pink-to-red colour indicated the presence of flavanoids¹³.

- c) **Test for saponins:** To 1 ml of the extract, 5 ml of distilled water was added and kept for few minutes. Frothing persistence indicated the presence of saponins¹³.
- d) **Test for alkaloids:** To about 3 ml of extract, a few drops of Mayer's reagent [prepared by dissolving 1.36g of mercuric chloride and 5g of potassium iodide in 100ml of water] were added along the sides of the test tube. Development of cream coloured precipitate inferred the presence of alkaloids¹⁴.
- e) **Test for tannins:** About 200 mg of flower material was extracted with distilled water and then filtered. To the filtrate, drops 0.1 % of FeCl₃ was added. A blue-black colouration, indicated the presence of tannins¹⁴.
- f) **Test for quinones:** To 1 ml of the ethanolic extract, 1 ml of concentrated sulphuric acid was added. Presence of quinones was indicated by development of red colour¹⁵.
- g) **Test for coumarins:** Above 1 ml of extract was mixed with a few drops of 10% sodium hydroxide. Development of yellow colour exhibited the presence of coumarins¹⁵.

GC-MS analysis

The GC-MS study was carried out using Clarus500 GC Perkin Elmer instrument in Food Testing Laboratory at Indian Institute of Crop Processing Technology, Thanjavur. The analysis was carried out for 72 minutes with Turbo mass detector¹⁴.

Antibacterial activity

Antibacterial activity of the ethanolic extract was tested against *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas putidaby* well diffusion method. The extract was injected into the well in different volume of 100µL, 200µL, 300µL and 400µL using sterile syringe. The plates were incubated at 37°C for 24 hours for bacterial growth. The plates were then observed for the zone of clearance around the well and measured in mm. The diameter of the inhibition zone was taken in four different fixed directions^{15, 16}.

RESULTS AND DISCUSSION

In the recent years, research on medicinal plants has attracted a lot of attention globally. Large body of evidence has accumulated to demonstrate the promising potential of medicinal plants used in various traditional, complementary and alternate systems of

treatment of human diseases. Plants are rich in a wide variety of secondary metabolites such as tannins, terpenoids, alkaloids and flavonoids etc, which have been found in vitro to have antibacterial and antifungal properties.^{17, 18}

Preliminary phytochemical screening of ethanolic extract

The phytoconstituents present in the ethanolic extract of *Terminalia catappa* flowers were identified by preliminary phytochemical screening and GC-MS analysis. The screening of ethanolic extract was carried out for different class of compounds and the results indicate the presence of *terpenoids*, *flavonoids* and *alkaloids* in the extract. The extraction and analysis of chemical constituents is an eco-friendly approach.^{19, 20}

Compounds identified from GCMS analysis

The ethanolic extract was subjected to GCMS analysis for the identification of compounds present in it. GCMS analysis shows that n-Hexadecanoic acid (33.67%), Tetradecanoic acid (16.37%) and 9, 12 – Octadecadienoic acid (14.5%) are present in higher level while the other volatile compounds like 2,6,6-Trimethyl-2,4-cycloheptadien-1-one, 4,6,6-Trimethylbicyclo(3.1.1)hept-3-en-2-one, 2,2-

dimethyl-3-(2-methylprop-1-enyl)cyclopropane-1-carboxylic acid, squalene and cedrane, 8-propoxy- are present in very low level.

The GCMS spectrum of ethanolic extract of *Terminalia catappa* indicating the presence of chemical constituents [Table 1] is given in Figure 1.

Antibacterial activity of ethanolic extract of *Terminalia catappa* flowers

The antibacterial activity of ethanolic extract was tested in different concentrations against *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas putida*. The zone of inhibition was measured after 24 hours and the results presented in Table 2.

Significant antibacterial activity was observed for 300 µL and 400 µL of ethanol extract against all the pathogens tested. Maximum inhibition activity was found for 400 µL with the inhibition zone of 8 mm against *Escherichia coli* followed by 7 mm against *Streptococcus aureus* [Figure 2]. The inhibition activity for 100 µL was minimum for the all the organism.

The phytochemical analysis reveals that the bioactive compounds n-hexadecanoic acid, tetradecanoic acid and 9, 12 – octadecadienoic acid in the extract are also responsible for antibacterial activity.

Table 1: Compounds identified from GCMS analysis

S.No	Compound	Molecular Formula	Molecular Weight in g/mol	Retention Time	Peak Area %
1.	2,6,6-Trimethyl-2,4-cycloheptadien-1-one	C ₁₀ H ₁₄ O	150.21	6.74	1.35
2.	4,6,6-Trimethylbicyclo[3.1.1]hept-3-en-2-one	C ₁₀ H ₁₄ O	150.22	6.92	1.13
3.	5-Hydroxymethyl-2-furaldehyde	C ₆ H ₆ O ₃	126.11	7.77	1.15
4.	2,2-dimethyl-3-(2-methylprop-1-enyl)cyclopropane-1-carboxylic acid	C ₁₀ H ₁₆ O ₂	168.232	10.05	6.56
5.	Dodecanoic acid [Lauric acid]	C ₁₂ H ₂₄ O ₂	200.32	11.00	2.38
6.	Tetradecanoic acid [Myristic acid]	C ₁₄ H ₂₈ O ₂	228.37	13.42	16.34
7.	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256.43	16.23	33.67
8.	Ethyl hexadecanoate [Ethyl palmitate]	C ₁₈ H ₃₆ O ₂	284.48	16.46	4.04
9.	Tetradecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethylester [Synonyms: Myristic acid β-monoglyceride]	C ₁₇ H ₃₄ O ₄	302.45	17.22	0.46
10.	9,12-Octadecadienoic acid(Z,Z)-	C ₁₈ H ₃₂ O ₂	280.44	18.72	14.54
11.	Octadecanoic acid [Stearic acid]	C ₁₈ H ₃₆ O ₂	284.48	19.17	7.80
12.	1,2- Benzenedicarboxylic acid, diisooctyl ester	C ₂₄ H ₃₈ O ₄	390.55	24.66	1.68
13.	3,5,5-trimethyl-4-(3-oxobutyl)cyclohex-2-en-1-one	C ₁₃ H ₂₀ O ₂	208.29	27.64	3.35
14.	Squalene	C ₃₀ H ₅₀	410.73	28.83	3.79
15.	8-propoxycedrane,	C ₁₈ H ₃₂ O	264.44	31.35	1.73

Table 2: Antibacterial activity of ethanolic extract of *Terminalia catappa* flowers

Volume of ethanolic extract per well (µl)	Diameter of Inhibition zone (mm)		
	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas putida</i>
100	2	3	4
200	4	5	5
300	5	6	5
400	7	8	5

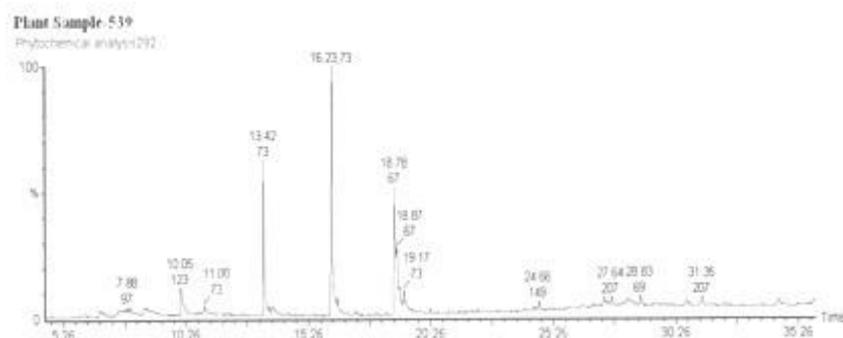


Fig. 1: GCMS spectrum of the ethanolic extract of the *Terminaliacatappa* flower

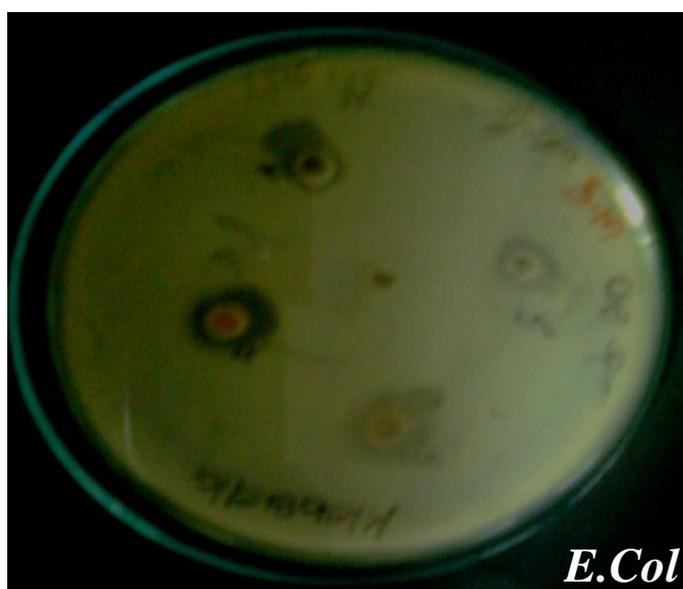


Fig. 2: Zone of Inhibition for ethanolic extract against *Escherichia coli*

CONCLUSION

Terminaliacatappa is an important tropical plant grown in many parts of Tamil Nadu, India. The *Terminaliacatappa* possess good nutritional, biological and medicinal value.

In the present study the ethanolic extract of *Terminaliacatappa* flowers was prepared and it was found to

1. Contain many organic compounds which cannot be easily synthesized.
2. Have good antibacterial activity. The phytochemicals present in the *Terminaliacatappa* flower contribute for its antibacterial properties.

ACKNOWLEDGEMENT

The Authors thank the Principals and Managements of Bishop Heber College (Autonomous), Tiruchirappalli, Government Arts College, Ariyalur and Syed Ammal Engineering

College, Ramanathapuram Tamil Nadu, India for their encouragement and support.

REFERENCES

1. Shikha Mandloi, Renu Mishra, Ranjana Verma, Shubhangi Mugal and Rajshree S. Phytochemical analysis of the leaf extract of *Terminaliacatappa* L. *Indian J. Applied & Pure Bio.* 2013; 28(1): 65-70
2. Vipul R. Thummar, Subramani Parasuraman, Debdatta Basu and Ramasamy Raveendran. Evaluation of in vivo antitumor activity of cleistanthin B in Swiss albino mice. *Journal of Traditional and Complementary Medicine.* 2015; 1-6
3. Ahmad I, Mehmood Z and Mohammad F. Screening of some

- Indian medicinal plants for their antimicrobial properties. *Journal of Ethnopharmacol.* 1998; 62(2): 183-193
4. AtesDA and OzlemTurgayErdogru. Antimicrobial activities of various medicinal and commercial plant extracts. *Turkish Journal of Biology.* 2003; 27(3):157-162
 5. 5.NairR and SumitraChanda. Antimicrobial activity of Terminaliacatappa, Manilkarazapota and Piper betel leaf extract. *Indian Journal of Pharmaceutical Sciences.* 2008 May-Jun; 70(3): 390-393.
 6. 6.Akharaiyi FC, Ilori RM and AdesidaJA. Antibacterial effect of Terminaliacatappa on some selected pathogenic bacteria. *Int J Pharm Biomed Res.* 2011; 2(2): 64-67
 7. 7.Parekh J and Chanda SV. In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turkish Journal of Biology.* 2007; 31(1): 53-58
 8. 8.ArunachalamP, SankarM and Subramanian B. Antibacterial activity of plant extract against plant bacterial pathogens. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences.* 2010; 12(1): 167-170
 9. 9.Arumugam VijayaAnand, NatarajanDivya, Panner selvamPunniyaKotti. An updated review of Terminaliacatappa. *Pharmacognosy Review.* 2015; 9(18):93-98
 10. Syed Mansoor Ahmed, VrushabendraSwamy BM, Gopkumar R, Dhanapal P and ChandrashekarVM. Anti-Diabetic activity of Terminaliacatappa Linn. Leaf extracts in alloxan-induced diabetic rats. *Iranian Journal of Pharmacology and Therapeutics.* 2005; 4:36-39
 11. Matos L, Nzikou JM, KimbonguilaA, Ndangui CB, Pambou-Tobi NPG, Abena AA and Desorby S. Composition and nutritional properties of seeds and oil from Terminaliacatappa L. *Advance Journal of Food Science and Technology.* 2009; 1(1): 72-77
 12. 12.Neelavathi P, Venkatalakshmi P and Brindha P. Antibacterial activities of aqueous and ethanolic extracts of Terminaliacatappa leaves and bark against some pathogenic bacteria. *International Journal of Pharmacy and Pharmaceutical Sciences.* 2013; 1(1): 114-120
 13. Prakash G and HosettiBB. Bio-efficacy of *Dioscorea pentaphylla* from Mid-Western Ghats, India. *Toxicology International.* 2012; 19 (2): 100-105
 14. 14.Ayoola GA, Coker HAB, Adesegun SA, Adepoju-BelloAA, ObaweyaK, EzenniaEC and AtangbayilTO. Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria therapy in Southwestern Nigeria. *Tropical Journal of Pharmaceutical Research.* 2008; 7(3):1019-1024
 15. 15. SeemaFirdouse and ParwezAlam. Phytochemical investigation of extract of *Amorphophallus campanulatus* tubers. *International Journal of Phytomedicine.* 2011; 3(1): 32-35
 16. 16. Moses S, Owolabi, Oladipupo A, Lawal, Isiaka A, Ogunwande, Rebecca M, Hauser and William N Setzer. Chemical composition of the leaf essential oil of Terminaliacatappa L. growing in Southwestern Nigeria. *American Journal of Essential Oils and Natural Products.* 2013; 1 (1): 51-54
 17. 17. Sher A. Analysis of typhoid ileal perforation. *Gomal Journal of Medical sciences.* 2009; 7(1):72-78
 18. 18. Timothy SY, Wazis CH, Bwala AY, Bashir HJ, Rhoda AS. Comparative study on the effects of aqueous and ethanol leaf extracts of cassia alata Linn on some pathogenic bacteria and fungi. *International Research Journal of pharmacy.* 2012; 3(8):125-127
 19. 19. Krishnaveni M, Krishna Kumari G, Kalaivani M, Ragina Banu C. Gas chromatography – mass spectrometry/mass spectrometry analysis of Terminaliacatappa L. nut and antimicrobial assay. *Asian journal of Pharmaceutical and Clinical Research.* 2015; 8(4):168-170
 20. 20. Offor CE, Ugwu PC, Okechukwu, Aja PM, and Igwenyi IO. Proximate and phytochemical analyses of Terminaliacatappa leaves. *European Journal of Applied Sciences* 2015; 7 (1): 09-11